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Syntheses of model compounds related to an antigenic epitope in pectic polysaccharides from *Bupleurum* falcatum L.

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Abstract

Stereocontrolled syntheses of model compounds related to a category of the major antigenic epitope against anti-bupleurum 2IIc/PG-1-IgG from an anti-ulcer pectic polysaccharide are described. Glycosylation of the glucuronic acid donors methyl(2,3-di-O-benzoyl-4-O-methyl- α -D-glucopyranosyl trichloroacetimidate)uronate and methyl (2,3-di-O-benzoyl-4-O-methyl- β -D-glucopyranosyl)uronate-(1 \rightarrow 6)-2,3,4-tri-O-benzoyl- α -D-galactopyranosyl trichloroacetimidate with the common acceptor 2-(trimethylsilyl)ethyl 2,3,4-tri-O-benzyl- β -D-galactopyranoside in the presence of trimethylsilyl triflate (TMSOTf) gave the desired di- and trisaccharide derivatives. Furthermore the products were transformed into the oligo-valent clustering saccharides, N,N',N''-tri- $\{5$ -[4-O-methyl- β -D-galactopyranosyloxy]pentylcarbonylaminoethyl $\}$ -1,3,5-benzenetriamide and N,N',N''-tri- $\{5$ -[4-O-methyl- β -D-galactopyranosyluronic acid (1 \rightarrow 6)- β -D-galactopyranosyloxy]pentylcarbonylaminoethyl $\}$ -1,3,5-benzenetriamide. © 2001 Elsevier Science Ltd. All rights reserved.

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1. Introduction

In the last few years, it has become apparent that pectic polysaccharides including pectin from medicinal herbs display various in vitro and in vivo pharmacological activities in addition to the application for drug delivery. Therefore, it is possible to consider pectic polysaccharides, not only as one of the active ingredients of medicinal herbs, but also as medicinals.¹ Recently, Yamada et al.² have

reported that the potent anti-ulcer pectic polysaccharide (Bupleuran 2IIc) was isolated from the hot-water extract of the roots of *Bupleurum falcatum*. Bupleuran 2IIc consists of a galacturonan region, a 'ramified' region (PG-1) composed of a rhamnogalacturonan core having neutral sugar side chains, and a rhamnogalacturonan II-like region;³ the 'ramified' region has been considered to be an important part of the expression of the activity. The backbone of PG-1 is composed of an α -L-Rha- $(1 \rightarrow 4)$ - α -D-GalA- $(1 \rightarrow 2)$ -repeating unit, which was reported in our previous paper⁴ as the synthetic tetrasaccharide.

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On the other hand, a polyclonal antibody (anti-bupleuran 2IIc/PG-1-IgG) against the PG-1 of the bupleuran 2IIc was prepared, and the major antigenic epitope against this antibody was characterized as a 6-linked galactosyl chain containing terminal glucuronic acid 4-*O*-methyl-glucuronic or (GlcA4Me), which were substituted to $(1 \rightarrow 3)$ β-D-galactosyl chains in the 'ramified' region of bupleuran 2IIc.5 Furthermore, among the structural region of bupleuran 2IIc, the 'ramified' region (PG-1), showed potent mitogenic activity, suggesting it was the active site.⁶ The proposed structure of the antigenic epitope in PG-1 was the target for synthetic studies as part of our investigations on the synthesis of oligosaccharides of biological interest. In this time, di-, and trisaccharide derivatives including GlcA4Me were chosen as target compounds. Concerning the synthesis of oligosaccharides, suitable glycomimetics that are able to compete or perform even better than the naturally occurring carbohydrate ligands are needed for the development of suitably bioactive compounds. For this purpose clustering glycosides⁷ have proved to be advantageous in many instances, as the multipresentation of specific sugar epitope in one molecule can result in remarkably increased adhesion. It is expected that multivalent saccharides would bind to the cell-surface adhesion molecule more tightly than monovalent ones.

We therefore initiated the synthesis of simple readily accessible oligo-valent saccharides, and we report herein the synthesis of monoand trivalent analogs (1–4) of β -D-GlcA4Me-(1 \rightarrow 6)- β -D-Gal- and β -D-GlcA4Me-(1 \rightarrow 6)- β -D-Gal-(1 \rightarrow 6)- β -D-Gal.

2. Results and discussion

Syntheses of monosaccharide derivatives.— Syntheses of the additional glucuronic acid methyl ester building blocks **9** were carried out as depicted in Scheme 1. Compound **6** was prepared from known 2-(trimethylsilyl)ethyl 2,3-di-*O*-benzyl-β-D-glucopyranoside (**5**)⁸ by the following three-step procedure. Regioselective tritylation of the starting material with trityl chloride, followed by methylation and subsequent acid-hydrolysis of the trityl group, gave compound **6**. Oxidation and esterifica-

tion of **6** was carried out with K₂Cr₂O₇ and HCl–MeOH. Removal of the benzyl groups from **7** by catalytic hydrogenolysis over 10% Pd–C and subsequent benzoylation gave methyl[2 - (trimethylsilyl)ethyl(2,3 - di - O - benzoyl-4-O-methyl-β-D-glucopyranosid)uronate)] (**8**). For selective removal of the 2-(trimethylsilyl)ethyl (SE) group, **8** was treated⁹ with trifluoroacetic acid in dichloromethane for 1 h at 0 °C to give the 1-hydroxy compound, which on further treatment¹⁰ with trichloroacetonitrile in the presence of 1,8-diazabicyclo[5.4.0]-

undec-7-ene (DBU) in dichloromethane for 2 h at 0 °C, gave the corresponding α -trichloroacetimidate **9** (70%) as the sole product (Scheme 1).

Syntheses of target oligosaccharide analogues.—The glycosylation of **9** with 2-(trimethylsilyl)ethyl 2,3,4-tri-*O*-benzyl-β-D-galactopyranoside (**10**) in the presence of trimethylsilyl triflate (TMSOTf) and 4 Å MS in dichloromethane for 1 h at 0 °C gave the desired disaccharide **11** (81%), as evidenced by ¹H NMR spectroscopy (H-1′, 4.81 ppm, *J* 7.9

Scheme 2.

Hz). Compound 11 was subjected to O-debenzylation, followed by benzoylation, to give compound 12. Selective removal of the SE group and treatment with trichloroacetonitrile in the presence of DBU gave only the corresponding α-trichloroacetimidate 13. Glycosylation of 13 with 10 in the presence of TMSOTf and 4 Å MS in dichloromethane for 1 h at 0 °C gave the desired trisaccharide 14 (85%), as evidenced by ¹H NMR spectroscopy (H-1', 4.80 ppm, J 7.9 Hz), showing the newly formed glycosidic linkages to be β. Compound 14 was converted by O-debenzylation and subsequent benzoylation to compound 15. Selective removal of the SE group, and treatment with trichloroacetonitrile in the presence of DBU gave only the corresponding αtrichloroacetimidate 16. In an alternate procedure, a part of 12 and 15 was converted by debenzoylation and hydrolysis of the ester to give the target compounds 1 and 2 (Scheme 2).

Next, concerning the synthesis of sugar clusters, spacer groups are needed as linkers between the core and the carbohydrate chain.

We chose 5-[2-(2,2,2-trichloroethoxycarbonyl)aminoethyleneaminocarbonyl]pentanol (20) as a spacer, which was prepared as depicted in Scheme 3. Compound 20 was prepared from \(\varepsilon\)-caprolactone by the following procedure. E-Caprolactone was converted to 5-methoxycarbonylpentanol by NaOMe, and subsequent benzylation gave benzyl methoxycarbonylpentyl ether (17). Compound 17 was converted to the primary amine 18 by reaction with neat anhydrous ethylenediamine¹¹ and was protected by 2,2,2-trichloroethyl carbamate (19). Finally removal of the benzyl groups by catalytic hydrogenolysis over 10% Pd-C gave the target spacer, 5 - [2 - (2.2.2 - trichloroethoxycarbonyl)aminoethyleneaminocarbonyl]pentanol (TOF-MS m/z: 371.3 [M + Na]⁺) (20) (Scheme 3). Coupling of 13 and/or 16 with the spacer 20 in the presence of TMSOTf and 4 Å MS for 1 h at 0 °C afforded the desired di- and trisaccharide derivatives 21 (65%) and 22 (60%), as evidenced by ¹H NMR spectroscopy (21: H-1, 4.58 ppm, J 7.9 Hz, **22**: H-1, 4.68 ppm, J 7.9 Hz). Removal of the Troc group from 21 and

Scheme 3.

Scheme 4.

22 by Zn-AcOH gave the primary amines 23 and 24 (Scheme 4). We chose 1,3,5-benzenetricarbonyl trichioride¹² a core for the synthesis of the sugar cluster. Coupling of 23 and/or 24 1,3,5-benzenetricarbonyl with core, trichloride in the presence of triethylamine afforded the desired trivalent oligosaccharide cluster **25** (64%) and **26** (78%), as evidenced by TOF-MS (25: $[M + Na]^+ = 3360$, 26: $[M + Na]^+ = 3360$ $Na]^+ = 4782$) (Scheme 5). Finally, removal of all acyl groups and esters with sodium methoxide in 5:1 methanol-water for 4 h at room temperature afforded the desired oligosaccharide clusters 3 and 4. Compound 3 revealed an $[M + Na]^+$ ion peak at m/z 1757 and compound 4 revealed an $[M + Na]^+$ ion peak at m/z 2243 in the TOF-MS spectrum.

Mitogenic activity of these compounds on spleen cells and Peyer's patch cells were evaluated, and the results will be reported in detail elsewhere.

3. Experimental

Optical rotations were determined with a JASCO digital polarimeter. ¹H and ¹³C NMR spectra were recorded on a JNM A 500 FT NMR spectrometer in CDCl₃ with Me₄Si as the internal standard. MALDI-TOFMS was recorded on a Perceptive Voyager RP mass spectrometer. TLC was performed on Silica Gel 60 F₂₅₄ (E. Merck) with detection by quenching of UV fluorescence and by spraying with 10% H₂SO₄. Column chromatography was carried out on Silica Gel 60 (E. Merck). 2-(Trimethylsilyl)ethyl 2,3-di-*O*-benzyl-β-D-glucopyranoside (5) was prepared by a literature method.⁸

2-(Trimethylsilyl)ethyl 2,3-di-O-benzyl-4-O-methyl-β-D-glucopyranoside (6).—To a solution of 2-(trimethylsilyl)ethyl 2,3-di-O-benzyl-β-D-glucopyranoside (5) (1.7 g, 3.93 mmol) in pyridine (15 mL) was added triphenylmethyl chloride (2.2 g, 7.89 mmol). The reaction mixture was stirred at 70 °C for 12 h, then diluted with CHCl₃ and washed with 5% HCl, aq NaHCO₃ and water, dried (Na₂SO₄), and concentrated. The product was purified by silicagel column chromatography using 100:1 benzene–acetone as eluent to give 2-(tri-

methylsilyl)ethyl 2,3-di-O-benzyl-6-O-trityl-β-D-glucopyranoside. To a solution of this compound in DMF (15 mL) was added NaH in oil (110 mg) and iodomethane (1 mL). The mixture was stirred for 4 h at rt, and then MeOH was added to destroy excess NaH. The reaction mixture was poured into ice-water and extracted with CHCl₃. The extract was successively washed with aq NaHCO₃ and water, dried (Na₂SO₄), and concentrated. This compound was treated with 80% AcOH (10 mL) at 55 °C for 10 h, and following removal of AcOH and water by coevaporation with toluene, the crude product was purified by column chromatography (50:1 benzene-acetone) on silica gel to give compound 6 (1.31 g, three steps 70%): ¹H NMR (CDCl₃): δ 7.34– 7.25 (m, 10 H, $2 \times Ph$), 4.95-4.69 (m, 4 H, 2 $PhCH_2$), 4.42 (d, 1 H, $J_{1.2}$ 7.9 Hz, H-1), 4.00- $3.95 \text{ (m, 1 H, } -OCH_2CH_2-), } 3.93-3.76 \text{ (br,d, 2)}$ H, H-6a,6b), 3.63-3.26 (m, 5 H, H-2,3,4,5, $-OCH_2CH_2-$), 3.55 (s, 3 H, OMe), 0.97-0.86 (m, 2 H, $-OCH_2CH_2$), -0.06 (s, 9 H, MALDI-TOFMS: $Si(CH_3)_3$: Calcd for $C_{26}H_{38}O_6Si: m/z$ 474.2. Found: m/z 497.2 $[M + Na]^+$.

Methyl [2-(trimethylsilyl)ethyl 2,3-di-O-ben $zvl - 4 - O - methvl - \beta - D - glucopyranosidluronate$ (7).—To a solution of **6** (1.22 g, 5.06 mmol) in acetone (30 mL) was added K₂Cr₂O₇ (2.8 g), 3.5 M $H_2SO_4(11 \text{ mL})$, and the mixture was stirred for 1 h at 55 °C, at the end of which time water was added. The residue was washed with CHCl3, and the washings were successively washed with water. dried (Na₂SO₄), and concentrated. The resulting compound was treated with 10% HCl-MeOH (60 mL) at rt for 2 h and then concentrated. The product was purified by silica-gel column chromatography using 8:1 hexane-EtOAc as eluent to give 7 (1.88 g, 73.9%): ¹H NMR $(CDCl_3)$: δ 7.36–7.26 (m, 10 H, 2 Ph), 4.94– 4.68 (m, 4 H, 2 PhC H_2), 4.42 (d, 1 H, J_1 , 7.5 Hz, H-1), 4.00-3.94 (m, 1 H, $-OCH_2CH_2-$), 3.81 (s, 3 H, COOMe), 3.78 (d, 1 H, $J_{4.5}$ 10.5 Hz, H-5), 3.62-3.39 (m, 4 H, H-2,3,4, $-OCH_2CH_2-$), 3.49 (s, 3 H, OMe), 0.96-0.86 (m, 2 H, $-OCH_2CH_2$), -0.03 (s, 9 H, $Si(CH_3)_3$); ¹³C NMR (CDCl₃): δ 169.1 (C-6), 103.4 (C-1), 83.7 (C-4), 81.6 (C-2), 81.1 (C-3), 75.5, 74.8 (2 Ph CH_2), 74.3 (C-5), 67.8

($-OCH_2CH_2-$), 60.6 (COOMe), 52.4 (OMe), 18.4 ($-OCH_2CH_2-$), -1.5 ($SiMe_3$). MALDITOFMS: Calcd for $C_{27}H_{38}O_7Si$: m/z 502.2. Found: m/z 525.1 [M + Na] + .

Methyl [2-(trimethylsilyl)ethyl 2,3-di-O-ben $zoyl-4-O-methyl-\beta-D-glucopyranosid$ furonate (8).—A solution of 7 (1.87 g, 3.73 mmol) in MeOH (10 mL) and THF (10 mL) was hydrogenated over 10% Pd-C (230 mg) for 3 h at rt, filtered through Celite, and the residue was washed with MeOH and concentrated. The residue was benzovlated with BzCl (4 mL) in pyridine (15 mL) for 10 h at rt. The reaction mixture was poured into ice-water and extracted with CHCl₃. The extract was washed sequentially with 5% HCl, aq NaHCO₃ and water, dried (Na₂SO₄), and concentrated. The product was purified by silica-gel column chromatography using 5:1 hexane-EtOAc as eluent to give **8** (1.84 g, 93.1%): 1 H NMR: δ 8.07-7.34 (10 H, m, 2 Ph), 5.68 (t, 1 H, $J_{2.3} = J_{3.4}$ 9.2 Hz, H-3), 5.43 (t, 1 H, $J_{1.2}$ 7.6 Hz, H-2), 4.82 (d, 1 H, H-1), 4.15 (d, 1 H, $J_{4.5}$ 9.6 Hz, H-5), 4.12-3.97 (m, 2 H, H-4, $-OCH_2CH_2-$), 3.92 (s, 3 H, COOMe), 3.69-3.50 (m, 1 H, $-OCH_2CH_2$), 3.48 (s, 3 H, OMe), 0.92-0.87 (m, 2 H, $-OCH_2CH_2$), -0.03 (s, 9 H, Si(CH₂)₂); 13 C NMR (CDCl₂): δ 168.6 (C-6), 100.8 (C-1), 79.1 (C-4), 74.4 (C-3,5), 72.0 (C-2), 67.8 ($-OCH_2CH_2-$), 60.4 (COOMe), 52.7 (OMe), 17.8 $(-OCH_2CH_2-)$, -1.5 (SiMe₃). MALDI-TOFMS: Calcd for $C_{27}H_{24}O_{0}Si: m/z$ 530.2. Found: m/z 553.3 $[M + Na]^+$.

Methyl (2,3-di-O-benzovl-4-O-methyl- α -Dtrichlornacetimidate)uronate glucopyranosyl (9).—To a solution of **8** (115 mg, 0.22 mmol) in CH₂Cl₂ (2 mL) cooled to 0 °C was added CF₃COOH (1 mL), and the mixture was stirred for 1 h at 0 °C and concentrated. Ethyl acetate and toluene (1:2) were added, and the solvent was evaporated to give the 1-hydroxy compound. To a solution of the residue in CH₂Cl₂ (1 mL), cooled at 0 °C were added trichloroacetonitrile (0.7 mL, 7 mmol) and DBU (60 µL, 0.4 mmol). The mixture was stirred for 2 h at 0 °C. After completion of the reaction, the mixture was concentrated. chromatography Column (3:1)hexane-EtOAc) of the residue on silica gel gave 9 (89 mg, 70%) as an amorphous mass: ¹H NMR

(CDCl₃): δ 8.63 (s, 1 H, NH), 8.04–7.26 (m, 10 H, 2 Ph), 6.76 (d, 1 H, $J_{1,2}$ 3.3 Hz, H-1), 6.07 (t, 1 H, $J_{2,3} = J_{3,4}$ 9.5 Hz, H-3), 5.46 (dd, 1 H, H-2), 4.54 (d, 1 H, $J_{4,5}$ 99 Hz, H-S), 3.98 (t, 1 H, H-4), 3.84 (s, 3 H, COOMe), 3.45 (s, 3 H, OMe); MALDI-TOFMS: Calcd for $C_{24}H_{22}Cl_3NO_9$: m/z 573.0. Found: m/z 596.1 [M + Na]⁺.

2-(Trimethylsilyl)ethyl [methyl (2,3-di-Obenzoyl - 4 - O - methyl - β - D - glucopyranosyl)*uronate*] - (1 → 6) - 2,3,4 - *tri* - O - *benzyl* - β - Dgalactopyranoside (11).—To a solution of the trichloroacetimidate 9 (636 mg, 1.11 mmol) and 10 (300 mg, 0.55 mmol) in CH₂Cl₂ (2 mL) were added 4 Å MS (500 mg), and the mixture was stirred for 3 h at rt, then cooled to 0 °C. TMSOTf (32 µL, 160 µmol) was added, and the mixture was stirred for 1 h at 0 °C, then neutralized with Et₃N. The solids were filtered off and washed with CHCl₃. The combined filtrate and washings were successively washed with water, dried (Na₂SO₄), and concentrated. The product was purified by silica-gel column chromatography using 2:1 hexane-EtOAc as eluent to give 11 (429 mg, 80.8%): $[\alpha]_{D}^{24}$ + 21.6° (c 1.1, CHCl₃). ¹H NMR (CDCl₃): δ 7.98–7.18 (m, 25 H, 5 Ph), 5.63 (t, 1 H, H-3'), 5.35 (t, 1 H, H-2'), 4.88-4.51 (m, 6 H, $3 \times$ CH_2Ph), 4.81 (d, 1 H, $J_{1'.2'}$ 7.9 Hz, H-1'), 4.24 $(d, 1 H, J_1, 7.9 Hz, H-1), 4.05 (d, 1 H, H-5'),$ 3.97, 3.75 (dt, 2 H, H-6), 3.94 (m, 1 H, H-4'), 3.87, 3.46 (dt, 2 H, $CH_2CH_2SiMe_3$), 3.78 (s, 3 H, COOMe), 3.77 (d, 1 H, H-4), 3.75 (m, 1 H, H-5), 3.72 (t, 1 H, H-2), 3.40 (s, 3 H, OMe), 3.35, 3.34 (dd, 1 H, H-3), 0.96 (t, 2 H, CH_2SiMe_3), -0.01 (s, 9 H, $SiMe_3$); ¹³C NMR $(CDCl_3)$: δ 168.4 (C-6'), 165.5, 165.1 (2 C=0), 103.3 (C-1), 101.2 (C-1'), 82.0 (C-3), 79.3 (C-2), 79.1 (C-4'), 75.1 (C-4, 2PhCH₂), 74.6 (C-5, $PhCH_2$), 74.3 (C-3'), 74.1 (C-5'), 72.0 (C-2'), 67.3 (-OCH₂CH₂-),67.9 (C-6),(COOMe), 52.7 (OMe), 18.4 (-OCH₂CH₂-), -1.1 (SiMe₃) MALDI-TOFMS: Calcd for $C_{54}H_{62}O_{14}Si: m/z 962.2.$ Found: m/z 985.3 $[M + Na]^+$.

2-(Trimethylsilyl)ethyl [methyl (2,3-di-Obenzoyl - 4 - O - methyl - β - D - glucopyranosyl)-uronate] - (1 \rightarrow 6) - 2,3,4 - tri - O - benzoyl - β - D-galactopyranoside (12).—A solution of 11 (121 mg, 0.13 mmol) in MeOH (4 mL) and THF (2 mL) was hydrogenated over 10%

Pd-C (100 mg) for 30 min at rt, then filtered through Celite. The filtrate was concentrated, and the residue was benzovlated with BzCl (0.5 mL) in pyridine (1.5 mL) for 1 h at rt. The reaction mixture was poured into ice-water and extracted with CHCl₃. The extract was washed sequentially with 5% HCl, NaHCO₃ and water, dried (Na₂SO₄), and concentrated. The product was purified by silicacolumn chromatography using benzene-EtOAc as eluent to give 12 (102 mg, 80.7%): $[\alpha]_D^{25} + 103.4^{\circ}$ (c 1.8 CHCl₃); ¹H NMR (CDCl₃): δ 8.06–7.19 (m, 25 H, 5 Ph), 5.84 (d, 1 H, H-4), 5.70 (t, 1 H, H-2), 5.62 (t, 1 H, H-3'), 5.48, 5.46 (dd, 1 H, H-3), 5.38 (t, 1 H, H-2'), 4.85 (d, 1 H, $J_{1'2'}$ 6.7 Hz, H-1'), 4.67 (d, 1 H, $J_{1.2}$ 8.6 Hz, H-1), 4.10 (m, 1 H, H-5), 4.06 (d, 1 H, H-5'), 4.05, 3.85 (dt, 2 H, H-6), 3.93 (t, 1 H, H-4'), 3.87, 3.44 (dt, 2 H, $CH_2CH_2SiMe_3$), 3.72 (s, 3 H, COOMe), 3.38 (s, 3 H, OMe), 0.73 (t, 2 H, CH_2SIMe_3), -0.12 (s, 9 H, SiMe₃); ¹³C NMR (CDCl₃): δ 168.4 (C-6'), 165.5, 165.2, 165.1 (5 C=O), 101.1 (C-1'), 100.8 (C-1), 78.8 (C-4'), 73.2 (C-5), 74.2 (C-3'), 74.2 (C-5'), 71.9 (C-2'), 71.9 (C-3), 69.9 (C-2), 69.9 (C-6), 68.8 (C-4), 67.5 $(-OCH_2CH_2-)$, 60.4 (COOMe), 52.7 (OMe), $17.7 \, (-OCH_2CH_2-), -1.4 \, (SiMe_2), MALDI-$ TOFMS: Calcd for $C_{54}H_{56}O_{17}Si: m/z$ 1004.3. Found: m/z 1027.4 [M + Na]⁺.

Methyl $(2,3-di-O-benzoyl-4-O-methyl-\beta-D-benzoyl-4-O-methyl-\beta-D-benzoyl-4-O-methyl-\beta-D-benzoyl-4-O-methyl-\beta-D-benzoyl-4-O-methyl-\beta-D-benzoyl-4-O-methyl-\beta-D-benzoyl-4-O-methyl-\beta-D-benzoyl-4-O-methyl-\beta-D-benzoyl-4-O-methyl-\beta-D-benzoyl-4-O-methyl-β-D-benzoyl-4$ glucopyranosyl)uronate - $(1 \rightarrow 6)$ - 2,3,4 - tri - Obenzoyl-\alpha-D-galactopyranosyl trichloroacetimidate (13).—To a solution of 12 (102 mg, 0.10 mmol) in CH₂Cl₂ (2 mL) cooled to 0 °C was added CF₃COOH (0.6 mL), and the mixture was stirred for 1 h at rt and concentrated. Ethyl acetate and toluene (1:2) were added, and the solvents were evaporated to give the 1-hydroxy compound. To a solution of the residue in CH₂Cl₂ (1 mL), cooled at 0 °C were added trichloroacetonitrile (0.4 mL, 4 mmol) and DBU (100 µL, 0.67 mmol). The mixture was stirred for 1 h at 0 °C. After completion of the reaction, the mixture was concentrated. Column chromatography (10:1 benzene-EtOAc) of the residue on silica gel gave 13 (87 mg, 83.0%) as an amorphous mass: ¹H NMR (CDCl₃): δ 8.32 (s, 1 H, NH), 8.08–7.22 (m, 25 H, 5 Ph), 6.73 (d, 1 H, $J_{1,2}$ 3.7 Hz, H-1), 6.00 (d, 1 H, H-4), 5.96 (m, 1 H, H-3), 5.84 (t,

1 H, H-2), 5.57 (t, 1 H, H-3'), 5.33 (t, 1 H, H-2'), 4.82 (d, 1 H, $J_{1',2'}$ 7.3 Hz, H-1'), 4.05 (m, 1 H, H-5), 4.05 (d, 1 H, H-5'), 4.05, 3.80 (dt, 2 H, H-6), 3.89 (t, 1 H, H-4'), 3.72 (s, 3 H, COOMe), 3.36 (s, 3 H, OMe)); 13 C NMR (CDCl₃): δ 168.4 (C-6'), 165.6, 165.4, 165.1, 160.4 (5 C=O), 100.9 (C-1'), 93.5 (C-1), 78.8 (C-4'), 74.2 (C-5), 74.2 (C-3'), 74.2 (C-5'), 71.7 (C-2'), 69.3 (C-3), 69.0 (C-2), 69.0 (C-4), 68.2 (C-6), 60.4 (COO*Me*), 52.7 (OMe). MALDITOFMS: Calcd for $C_{51}H_{44}Cl_3NO_{17}$: m/z 1048.6. Found: m/z 1071.6 [M + Na]⁺.

2-(Trimethylsilyl)ethyl [methyl (2,3-di-Obenzoyl - 4 - O - methyl - β - D - glucopyranosyl)*uronate*] - (1 → 6) - 2,3,4 - tri - O - benzoyl - β - Dgalactopyranosyl- $(1 \rightarrow 6)$ -2,3,4-tri-O-benzyl- β -D-galactopyranoside (14).—Compound 14 was prepared from 13 (700 mg, 0.67 mmol) and 10 (180 mg, 0.33 mmol) as described in the previous glycosylation procedure of disaccharide 11, yielding 474 mg (84.5%) of a syrup: $[\alpha]_D^{25} + 58.2^{\circ}$ (c 0.9 CHCl₃); ¹H NMR (CDCl₃): δ 4.80 (d, 1 H, $J_{1,2}$. 7.9 Hz, H-1'), 4.76 (d, 1 H, $J_{1',2'}$ 9.1 Hz, H- $\overline{1}''$), 4.29 (d, 1 H, $J_{1,2}$ 79 Hz, H-1), 3.55 (s, 3 H, COOMe), 3.39 (s, 3 H, OMe); 13 C NMR (CDCl₃): δ 104.8 (C-1), 102.6 (C-1'), 102.3 (C-1"). MALDI-TOFMS: Calcd for $C_{81}H_{84}O_{22}Si: m/z$ 1436.6. Found: m/z 1459.6 [M + Na)]⁺.

2-(Trimethylsilyl)ethyl [methyl (2,3-di-Obenzoyl - 4 - O - methyl - β - D - glucopyranosyl)-uronate] - (1 → 6) - 2,3,4 - tri - O - benzoyl - β - D-galactopyranosyl - (1 → 6) - 2,3,4 - tri - O - benzoyl-β-D-galactopyranoside (15).—Compound 15 was prepared from 14 (360 mg, 0.25 mmol) as described for the preparation of 13, yielding 307 mg (83.0%) of a syrup: $[\alpha]_{D}^{15}$ + 98.4° (c 0.7 CHCl₃); H NMR (CDCl₃): δ 4.81 (d, 1 H, $J_{1,2}$ 7.9 Hz, H-1), 4.74 (d, 1 H, $J_{1',2'}$ 7.9 Hz, H-1'), 4.61 (d, 1 H, $J_{1'',2''}$ 7.3 Hz, H-1''), 3.70 (s, 3 H, COOMe), 3.40 (s, 3 H, OMe). 13 C NMR (CDCl₃): δ 103.0 (C-1''), 102.9 (C-1'), 102.3 (C-1). MALDI-TOFMS: Calcd for $C_{81}H_{78}O_{25}$ Si: m/z 1479.6. Found: m/z 1502.6 [M + Na] +.

Methyl (2,3-di-O-benzoyl-4-O-methyl-β-D-glucopyranosyl)uronate - (1 \rightarrow 6) - 2,3,4 - tri - O-benzoyl-β-D-galactopyranosyl-(1 \rightarrow 6) - 2,3,4-tri-O-benzoyl-α-D-galactopyranosyl trichloro-acetimidate (16).—Compound 16 was prepared from 15 (100 mg, 67.6 μmol) as described for the preparation of 13, yielding 103 mg (quant) of a syrup: ¹H NMR (CDCl₃): δ

8.38 (s, 1 H, NH), 6.75 (d, 1 H, $J_{1,2}$ 3.7 Hz, H-1), 4.71 (d, 1 H, $J_{1',2'}$ 7.9 Hz, H-1'), 4.44 (d, 1 H, $J_{1'',2''}$ 7.3 Hz, H-1'), 3.62 (s, 3 H, COOMe), 3.36 (s, 3 H, OMe). ¹³C NMR (CDCl₃): δ 100.9 (C-1"), 100.7 (C-1'), 93.7 (C-1). MALDITOFMS: Calcd for $C_{78}H_{66}Cl_3NO_{25}$: m/z 1522.7. Found: m/z 1545.6 [M + Na]⁺.

2-(Trimethylsilyl)ethyl 4-O-methyl- β -Dglucopyranosyluronic acid- $(1 \rightarrow 6)$ - β -D-galactopyranoside (1).—To a solution of 12 (26 mg, 25.6 µmol) in 2:1 MeOH-water (3 mL) was added NaOMe (40 mg), and the mixture was stirred for 2 h at rt, then neutralized with Amberlite IR-120 (H⁺) resin. The resin was filtered off and washed with MeOH. The filtrate and washings were combined and concentrated. Column chromatography MeOH-water) of the residue on Sephadex LH-20 gave 1 (12.0 mg, quant): $[\alpha]_D^{25} - 43.5^{\circ}$ (c 0.6 MeOH); MALDI-TOFMS: Calcd for $C_{18}H_{34}O_{12}Si: m/z$ 470.5. Found: m/z 493.2 $[M + Na]^+$, 515.2 $[M + 2Na - H]^+$.

2-(Trimethylsilyl)ethyl 4-O-methyl- β -Dglucopyranosyluronic acid- $(1 \rightarrow 6)$ - β -D-galactopyranosyl - $(1 \rightarrow 6)$ - β - D - galactopyranoside (2).—To a solution of 15 (27 mg, 17.9 μmol) in 5:1 MeOH-water (2.4 mL) was added NaOMe (40 mg), and the mixture was stirred for 2 h at rt, then neutralized with Amberlite IR-120 (H⁺) resin. The resin was filtered off and washed with MeOH. The filtrate and washings were combined and concentrated. Column chromatography (3:1 MeOH-water) of the residue on Sephadex LH-20 gave 2 (11.3 mg, quant): $[\alpha]_D^{25} - 32.6^{\circ}$ (c 0.6 MeOH); MALDI-TOFMS: Calcd for $C_{24}H_{44}O_{17}Si: m/z$ 632.3. Found: m/z 655.3 [M + Na]⁺, 677.3 $[M + 2Na - H]^+$.

5-methoxycarbonylpentyl Benzyl ether (17).—To a solution of ε -caprolactone (10) mL, 90.2 mmol) in MeOH (100 mL) was added NaOMe (80 mg), and the mixture was stirred for 3 h at rt, then neutralized with Amberlite IR-120 (H⁺) resin. The resin was filtered off and washed with 1:1 CHCl₃-MeOH. The filtrate and washings were combined and concentrated. To a solution of this compound in DMF (262 mL) was added NaH in oil (19 g) and BnBr (56 mL). The mixture was stirred for 4 h at rt, and then MeOH was added to destroy excess NaH. The reaction mixture was poured into ice-water and extracted with CHCl₃. The extract was successively washed with aq NaHCO₃ and water, dried (Na₂SO₄), and concentrated. This compound was purified by column chromatography (5:1 hexane–EtOAc) on silica gel to give compound 17 (10.2 g, 47.7%): ¹H NMR (CDCl₃): δ 7.23–7.12 (m, 5 H, Ph), 4.44 (s, 2 H, CH₂Ph), 3.59 (s, 3 H, COOMe); MALDITOFMS: Calcd for C₁₄H₂₀O₃: m/z 236.1. Found: m/z 259.1 [M + Na]⁺.

5-(2-Aminoethyleneaminocarbonyl)pentyl benzyl ether (18).—Compound 17 (1 g, 4.5 mmol) was treated with ethylenediamine (10 mL) at 70 °C for 18 h. After completion of the reaction, the mixture was concentrated. Column chromatography (2:1 CHCl₃–MeOH) of the residue on silica gel to give 18 (917 mg, 77.1%) as an amorphous mass: 1 H NMR (CD₃OD): δ 7.38–7.32 (m, 5 H, aromatic), 4.48 (s, 2 H, CH₂Ph), 3.28 (br,s, 2 H, CH₂NHCOO), 2.79 (br,s, 2 H, CH₂NHCOO); MALDI-TOFMS: Calcd for C₁₅H₂₄N₂O₂: m/z 264.1. Found: m/z 287.1 [M + Na]⁺.

Benzyl 5-[2-(2,2,2-trichloroethoxycar-bonyl)aminoethyleneaminocarbonyl]pentyl ether (19).—To a solution of 18 (37 mg, 0.14 mmol) in pyridine (1 mL) cooled to 0 °C was added TrocCl (58 μL, 0.42 mmol), and the mixture was stirred for 30 min at rt and concentrated. Column chromatography (90:1 CHCl₃–MeOH) of the residue on silica gel gave 19 (50 mg, 80.3 %); ¹H NMR (CDCl₃): δ 7.34–7.27 (m, 5 H, Ph), 6.20 (br,s, 1 H, NHCO), 5.91 (br,s, 1 H, NHCOO), 4.71 (s, 2 H, CH₂CCl₃), 4.49 (s, 2 H, CH₂Ph), 3.36 (t, 2 H, CH₂NHCO); MALDI-TOFMS: Calcd for C₁₈H₂₅Cl₃N₂O₄: m/z 438.1. Found: m/z 461.0 [M + Na]⁺.

5-[2-(2,2,2-Trichloroethoxycarbonyl)amino-ethyleneaminocarbonyl]pentanol (20).—A solution of 19 (340 mg, 0.77 mmol) in MeOH (4 mL) was hydrogenated over 10% Pd–C (60 mg) for 20 min at rt, then filtered through Celite. The filtrate was concentrated, and the residue was purified by silica-gel column chromatography using 10:1 CHCl₃–MeOH as eluent to give 20 (178 mg, 65.8%); ¹H NMR (CDCl₃): δ 6.73 (br,s, 1 H, NHCO), 6.32 (br,s, 1 H, NHCOO), 4.72 (s, 2 H, CH₂CCl₃), 3.39 (t, 2 H, CH₂NHCO); MALDI-TOFMS: Calcd for C₁₁H₁₉Cl₃N₂O₄: m/z 348.0. Found: m/z 371.3 [M + Na]⁺.

5-[2-(2,2,2,-Trichloroethoxycarbonyl)aminoethyleneaminocarbonyl]pentyl [methyl (2,3-di-O-benzoyl-4-O-methyl- β -D-glucopyranosyl)uronate] - $(1 \rightarrow 6)$ - 2,3,4 - tri - O - benzoyl - β - Dgalactopyranoside (21).—To a solution of the trichloroacetimidate 13 (244 mg, 0.23 mmol) and **20** (40 mg, 0.12 mmol) in CH₂Cl₂ (1 mL) were added molecular sieves 4 Å (300 mg), and the mixture was stirred for 1 h at rt, then cooled to 0 °C. TMSOTf (22 µL, 120 µmol) was added, and the mixture was stirred for 1 h at 0 °C, then neutralized with Et₃N. The solids were filtered off and washed with CHCl₃. The combined filtrate and washings were successively washed with water, dried (Na₂SO₄), and concentrated. The product was purified by silica-gel column chromatography using 1:3 benzene-EtOAc as eluent to give 21 (100 mg, 64.7%): $[\alpha]_D^{24} + 74.2^{\circ}$ (c 0.7, CHCl₃). ¹H NMR (CDCl₃): δ 8.06–7.21 (m, 25 H, 5 × Ph), 5.80 (d, 1 H, H-4), 5.64 (t, 1 H, H-2), 5.61 (t, 1 H, H-3'), 5.48 (dd, 1 H, H-3), 5.37 (t, 1 H, H-2'), 4.85 (d, 1 H, $J_{1'2'}$ 7.3 Hz, H-1'), 4.69 (q, 2 H, OCH_2CCl_3), 4.58 (d, 1 H, J_1 , 7.9 Hz, H-1), 4.10 (m, 1 H, H-5), 4.07 (d, 1 H, H-5'), 4.03, 3.91 (dt, 2 H, H-6), 3.91 (t, 1 H, H-4'), 3.78 (s, 3 H, COOMe), 3.38 (s, 3 H, OMe), 3.36 (m, 2 H, OCH_2CH_2), 3.36 (m, 2 H, CH_2NHCO_1), 3.26 (m, 2 H, CH_2NHCOO), 1.89 (t, 2 H, CH₂CO), 1.44 (m, 2 H, CH₂CH₂CO), 1.14 (m, 2 H, OCH₂C H_2), 0.88 (t, 2 H, OCH₂CH₂C H_2); MALDI-TOFMS: Calcd for C₆₀H₆₁Cl₃N₂O₂₀: m/z 1235.7 Found: m/z 1258.4 [M + Na]⁺.

5-[2-(2,2,2,-Trichloroethoxycarbonyl)aminoethyleneaminocarbonyl]pentyl [methyl (2,3-di-O-benzoyl-4-O-methyl- β -D-glucopyranosyl)uronate] - $(1 \rightarrow 6)$ - 2,3,4 - tri - O - benzovl - β - D $galactopyranosyl-(1 \rightarrow 6)-2,3,4-tri-O-benzoyl$ β-D-galactopyranoside (22).—Compound 22 was prepared from 16 (86 mg, 56.5 µmol) and 20 (10 mg, 28.7 µmol) as described in the previous glycosylation procedure of 21, yielding 29 mg (59.7%) of a syrup: $[\alpha]_D^{25} + 72.1^{\circ}$ (c 1.3 CHCl₃); ¹H NMR (CDCl₃): δ 4.77 (d, 1 H, $J_{1'',2''}$ 7.9 Hz, H-1"), 4.76 (d, 1 H, $J_{1',2'}$ 7.9 Hz, H-1'), 4.68 (d, 1 H, $J_{1,2}$ 7.9 Hz, H-1), 3.72 (s, 3 H, COOMe), 3.41 (s, 3 H, OMe). ¹³C NMR $(CDCl_3)$: δ 101.6 (C-1), 101.5 (C-1"), 101.4 (C-1'); **MALDI-TOFMS:** Calcd $C_{87}H_{83}Cl_3N_2O_{28}$: m/z 1709.6. Found: m/z $1732.8 \text{ } [M + Na]^+.$

5-(2-Aminoethyleneaminocarbonyl)pentyl $(2,3-di\text{-O-}benzoyl\text{-}4\text{-O-}methyl\text{-}\beta\text{-D-}$ glucopyranosyl)uronate] - $(1 \rightarrow 6)$ - 2,3,4 - tri - Obenzoyl- β -D-galactopyranoside (23).—To solution of 21 (185 mg, 0.15 mmol) in AcOH (10 mL) was added zinc powder (200 mg), and the mixture was stirred for 12 h at rt. The solids were filtered off and washed with CHCl₃. The combined filtrate and washings were successively washed with water, dried (Na_2SO_4) , and concentrated. The product was purified by silica-gel column chromatography using 10:1 CHCl₃-MeOH as eluent to give 23 (92 mg, 58.0%): $[\alpha]_D^{25}$ + 74.8° (c 0.6 CHCl₃); ¹H NMR (CDCl₃): δ 8.06–7.21 (m, 25 H, 5 Ph), 5.99 (br,s, 1 H, NHCO), 5.80 (d, 1 H, H-4), 5.65 (t, 1 H, H-2), 5.61 (t, 1 H, H-3'), 5.49, 5.47 (dd, 1 H, H-3), 5.38 (t, 1 H, H-2'), 4.84 (d, 1 H, $J_{1'2'}$ 7.3 Hz, H-1'), 4.59 (d, 1 H, $J_{1,2}$ 7.9 Hz, H-1), 4.10 (m, 1 H, H-5), 4.06 (d, 1 H, H-5'), 4.07, 3.80 (m, 2 H, H-6), 3.92 (t, 1 H, H-4'), 3.75 (s, 3 H, COOMe), 3.38 (s, 3 H, OMe), 3.28 (m, 2 H, OCH_2CH_2), 3.28 (m, 2 H, CH_2NHCO_{-}), 2.83 (br,d, 2 H, CH_2NH_2), 2.02 (br,s, 2 H, NH₂), 1.88 (t, 2 H, CH₂CO), 1.43 (m, 2 H, CH_2CH_2CO), 1.32 (m, 2 H, OCH_2CH_2), 1.12 (t, 2 H, $OCH_2CH_2CH_2$); MALDI-TOFMS: Calcd for C₅₇H₆₀N₂O₁₈: m/z 1060.4. Found: m/z 1083.4 [M + Na]⁺. 5-(2-Aminoethyleneaminocarbonyl)pentyl

[methyl (2,3-di-O-methyl- β -D-glucopyranosyl)-uronate] - (1 \rightarrow 6) - 2,3,4 - tri - O - benzoyl - β - D-galactopyranosyl- (1 \rightarrow 6) - 2,3,4 - tri - O - benzoyl- β -D-galactopyranoside (24).—Compound 24 was prepared from 22 (56 mg) as described for the preparation of 23, yielding 46 mg (91.5%) of a syrup: [α]_D²⁵ + 73.3° (c 0.6 CHCl₃). ¹H NMR (CDCl₃): δ 4.78 (d, 1 H, $J_{1'',2''}$ 7.9 Hz, H-1''), 4.76 (d, 1 H, $J_{1',2'}$ 7.9 Hz, H-1''), 4.64 (d, 1 H, $J_{1,2}$ 7.3 Hz, H-1), 3.70 (s, 3 H, COOMe), 3.40 (s, 3 H, OMe). ¹³C NMR (CDCl₃): δ 101.7 (C-1), 101.5 (C-1'), 101.4 (C-1''); MALDI-TOFMS: Calcd for $C_{84}H_{82}N_2O_{26}$: m/z 1534.6. Found: m/z 1557.9 [M + Na]+.

N,N',N"-Tri-{5-[methyl (2,3-di-O-benzoyl-4-O-methyl- β -D-glucopyranosyl)uronate]- $(1\rightarrow 6)$ -2,3,4-tri-O-benzoyl- β -D-galactopyranosyloxypentylcarbonylaminoethyl} - 1,3,5-benzenetriamide (25).—To a solution of 23 (23 mg, 21.6 µmol) in CH₂Cl₂ (1 mL) were added triethylamine (3 µL) and 1,3,5-benzenetricarbonyl trichloride (1.3 mg, 5.0 µmol). The mix-

ture was stirred for 5 min at rt. After completion of the reaction, the mixture was concen-Column chromatography trated. CHCl₃-MeOH) of the residue on silica gel gave **25** (11 mg, 64.1%); ¹H NMR (CDCl₃): δ 5.80 (d, 3 H, $J_{3,4}$ 3.0 Hz, H-4), 5.61 (t, 6 H, J 9.1 Hz, H-2, H-3'), 5.50 (dd, 3 H, J_{34} 3.7, J_{23} 10.4 Hz, H-3), 5.36 (t, 3 H, H-2'), 4.85 (d, 3 H, $J_{1'2'}$ 7.3 Hz, H-1'), 4.57 (d, 3 H, J_{12} 7.9 Hz, H-1), 4.10 (m, 3 H, H-5), 4.07 (d, 3 H, H-5'), 4.03, 3.91 (dt, 6 H, H-6), 3.91 (t, 3 H, H-4'), 3.73 (s, 9 H, COOMe), 3.37 (s, 9 H, OMe); ¹³C NMR (CDCl₃): 168.5, 132.6–128.3, 101.3, 101.1, 78.9, 76.8, 74.1, 73.3, 71.9, 71.6, 70.0, 69.8, 68.8, 68.6, 60.4, 52.7, 29.7–11.5. MALDI-TOFMS: Calcd for $C_{180}H_{180}N_6O_{57}$: m/z 3337.1. Found: m/z 3360.2 [M + Na]⁺.

 $N,N',N''-Tri-\{5-[methyl\ (2,3-di-O-benzoyl 4 - O - methyl - \beta - D - glucopyranosyl)uronate]$ $(1 \rightarrow 6)$ -2,3,4-tri-O-benzoyl- β -D-galactopyrano $syl - (1 \rightarrow 6) - 2,3,4 - tri - O - \beta - D - galactopyrano$ syloxypentylcarbonylaminoethyl] - 1,3,5 - benzenetriamide (26).—To a solution of 24 (26) mg, 16.8 µmol) in CH₂Cl₂ (1 mL) were added triethylamine (2.4 µL) and 1,3,5-benzenetricarbonyl trichloride (1 mg, 3.9 µmol). The mixture was stirred for 5 min at rt. After completion of the reaction, the mixture was concentrated. Column chromatography (20:1 CHCl₃-MeOH) of the residue on silica gel gave **26** (8.6 mg, 77.8%): MALDI-TOFMS: Calcd for $C_{261}H_{246}N_6O_{81}$: m/z 4759.5. Found: m/z 4782.0 [M + Na]⁺.

N,N',N''-Tri- $\{5$ -[4-O-methyl- β -D-glucopyranosylurnnic acid- $(1 \rightarrow 6)$ - β -D-galactopyranosyloxy|pentylcarbonylaminoethyl} - 1,3,5 - benzenetriamide (3).—To a solution of 25 (9.4 mg, 2.8 µmol) in 5:1 MeOH–water (1.2 mL) was added NaOMe (39 mg), and the mixture was stirred for 4 h at rt, then neutralized with Amberlite IR-120 (H⁺) resin. The resin was filtered off and washed with MeOH. The filtrate and washings were combined and con-Column chromatography centrated. MeOH-water) of the residue on Sephadex LH-20 gave 3 (4.9 mg, quant.): $[\alpha]_D^{25} - \bar{1}7.8^{\circ}$ (c 0.2 water); MALDI-TOFMS: Calcd for $C_{72}H_{114}NO_{42}$: m/z 1734.5. Found: m/z 1757.0 $[M + Na]^+$, 1823.0 $[M - 3H + 4Na]^+$.

N,N',N''-Tri- $\{5-[4-O-methyl-\beta-D-glucopy-ranosyluronic acid <math>(1 \rightarrow 6)$ - β -D-galactopyrano-

syl- (1 → 6)- β-D-galactopyranosyloxy]pentyl-carbonylaminoethyl}-1,3,5-benzenetriamide (4). —To a solution of **26** (4.3 mg, 0.9 μmol) in 5:1 MeOH-water (0.6 mL) was added NaOMe (16 mg), and the mixture was stirred for 4 h at rt, then neutralized with Amberlite IR-120 (H⁺) resin. The resin was filtered off and washed with MeOH. The filtrate and washings were combinedand concentrated. Column chromatography (3:1 MeOH-water) of the residue on Sephadex LH-20 gave **4** (2.0 mg, quant) $[\alpha]_D^{25}$ – 23.2° (c 0.1 water); MALDITOFMS: Calcd for C₉₀H₁₄₄N₆O₅₇: m/z 2220.8. Found: m/z 2243.2 [M + Na]⁺, 2309.5 [M – 3H + 4Na]⁺.

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References

- Yamada, H. Pectin and Pectinases; Elsevier Science: Amsterdam, 1996; p. 173.
- Yamada, H.; Sun, X.-B.; Matsumoto, T.; Ra, K.-S.; Hirano, M.; Kiyohara, H. Planta Med. 1991, 57, 555–559.
- 3. Hirano, M.; Kiyohara, H.; Matsumoto, T.; Yamada, H. *Carbohydr. Res.* **1995**, *251*, 145–162.
- Maruyama, M.; Takeda, T.; Shimizu, N.; Hada, N.; Yamada, H. Carbohydr. Res. 2000, 325, 83–92.
- Sakurai, M.; Kiyohara, H.; Matsumoto, T.; Tsumuraya, Y.; Hashimoto, Y.; Yamada, H. Carbohydr. Res. 1998, 311, 219–229.
- Sakurai, M.; Kiyohara, H.; Yamada, H. *Immunology* 1999, 97, 540–547.
- (a) Zanini, D.; Roy, R. J. Am. Chem. Soc. 1997, 119, 2088–2095;
 - (b) Roy, R. Top. Curr. Chem. 1997, 187, 241-274;
 - (c) Ashton, P. R.; Hounsell, E. F.; Jayaraman, N.; Nilsen, T. M.; Spencer, N.; Stoddart, J. F.; Young, M. *J. Org. Chem.* **1998**, *63*, 3429–3437.
- 8. Wilstermann, M.; Balogh, J.; Magnusson, G. J. Org. Chem. 1997, 62, 3659–3665.
- Jansson, K.; Ahlfors, S.; Frejd, T.; Kihlberg, J.; Magnusson, G.; Dahmen, J.; Noori, G.; Stenvall, K. J. Org. Chem. 1988, 53, 5629–5647.
- 10. Schmidt, R. R.; Grundler, G. Synthesis 1981, 885-887.
- 11. Kamath, V. P.; Diedrich, P.; Hindsgaul, O. *Glycoconjugate J.* **1996**, *13*, 315–319.
- 12. Jiao, H.; Hindsgaul, O. J. Carbohydr. Chem. **1999**, 18, 499–513.